

## MODIFIED COUMARINS. 20. FUROCUMARIN DERIVATIVES OF CYTISINE

M. V. Veselovskaya,<sup>1</sup> M. M. Garazd,<sup>1</sup>  
V. I. Vinogradova,<sup>2</sup> and V. P. Khilya<sup>3</sup>

UDC 547.814.5

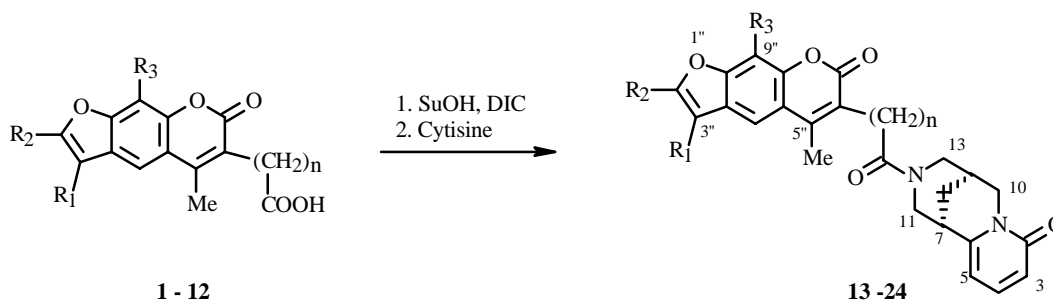
*Cytisine derivatives modified by furocoumarins were synthesized using activated esters.*

**Key words:** cytisine, coumarins, furocoumarins, acylation, activated esters.

Many important natural compounds are based on the furocoumarin structure [1]. Highly active pharmacological compounds are known among natural and synthetic furocoumarins [2].

On the other hand, the alkaloid cytisine and its derivatives possess a broad spectrum of biological activity [3, 4]. Therefore, a large number of publications on the chemical modification and study of the properties of cytisine have appeared [5-8]. Thus, addition to cytisine of a furocoumarin ring as a substituent is interesting both to the chemistry of alkaloids and coumarins and to the targeted search for new biologically active compounds.

Starting furocoumarins **1-12** that contain a carboxylic acid were prepared by the MacLeod method [9, 10]. Cytisine was *N*-acylated using activated esters [11], a method that is widely employed in peptide synthesis. The carboxylic acid was activated as the highly reactive *N*-hydroxysuccinimide ester [12]. Activated esters were prepared by reacting the corresponding acids **1-12** and *N*-hydroxysuccinimide (SuOH) in absolute dioxane using diisopropylcarbodiimide (DIC) as the condensing agent. Condensation of the resulting activated esters with cytisine in dioxane at room temperature gives in high yields the *N*-acyl cytisine derivatives **13-24**, the molecules of which contain furocoumarin moieties.



- 1, 13:** R<sub>1</sub> = Me, R<sub>2</sub> = R<sub>3</sub> = H, n = 1; **2, 14:** R<sub>1</sub> = R<sub>3</sub> = Me, R<sub>2</sub> = H, n = 1; **3, 15:** R<sub>1</sub> = R<sub>2</sub> = Me, R<sub>3</sub> = H, n = 1;  
**4, 16:** R<sub>1</sub> = R<sub>2</sub> = R<sub>3</sub> = Me, n = 1; **5, 17:** R<sub>1</sub>R<sub>2</sub> = (CH<sub>2</sub>)<sub>4</sub>, R<sub>3</sub> = H, n = 1; **6, 18:** R<sub>1</sub>R<sub>2</sub> = (CH<sub>2</sub>)<sub>4</sub>, R<sub>3</sub> = Me, n = 1;  
**7, 19:** R<sub>1</sub> = Me, R<sub>2</sub> = R<sub>3</sub> = H, n = 2; **8, 20:** R<sub>1</sub> = R<sub>3</sub> = Me, R<sub>2</sub> = H, n = 2; **9, 21:** R<sub>1</sub> = R<sub>2</sub> = Me, R<sub>3</sub> = H, n = 2;  
**10, 22:** R<sub>1</sub> = R<sub>2</sub> = R<sub>3</sub> = Me, n = 2; **11, 23:** R<sub>1</sub>R<sub>2</sub> = (CH<sub>2</sub>)<sub>4</sub>, R<sub>3</sub> = H, n = 2; **12, 24:** R<sub>1</sub>R<sub>2</sub> = (CH<sub>2</sub>)<sub>4</sub>, R<sub>3</sub> = Me, n = 2

1) Institute of Bioorganic Chemistry and Petroleum Chemistry, National Academy of Sciences of Ukraine, 02094, Ukraine, Kiev, ul. Murmanskaya, 1, e-mail: gmm@i.com.ua; 2) S. Yu. Yunusov Institute of the Chemistry of Plant Substances, Academy of Sciences of the Republic of Uzbekistan, Tashkent, fax (99871) 120 64 75; 3) Taras Shevchenko Kiev National University, 01033, Ukraine, Kiev, ul. Vladimirskaya, 64. Translated from *Khimiya Prirodnikh Soedinenii*, No. 3, pp. 230-232, May-June, 2006. Original article submitted November 16, 2005.

The PMR spectra revealed that a doubled set of signals was observed for all prepared compounds. Obviously invertomers with hindered rotation about the N–C bond that could be viewed as *Z*- and *E*-isomers were formed as a result of amide conjugation in these systems. The presence of amide conjugation was confirmed by variable temperature experiments. Thus, heating samples of the prepared compounds to 100°C caused the PMR signals to coalesce owing to free rotation around the N–C bond.

## EXPERIMENTAL

The course of reactions and purity of products were monitored by TLC on Merck 60 F254 plates using CHCl<sub>3</sub>:CH<sub>3</sub>OH (9:1 and 1:9) as eluent. Melting points were determined on a Kofler block. PMR spectra were recorded on a Varian VXR-300 spectrometer at 300 MHz relative to TMS (internal standard). Elemental analyses of all compounds agreed with those calculated.

Furocoumarins **1-12** were synthesized as before [9, 10]. We used pharmacopeic cytosine isolated from *Thermopsis lanceolata*.

**General Method of Cytosine *N*-acylation.** A solution of acids **1-12** (3 mmol) and *N*-hydroxysuccinimide (0.38 g, 3.3 mmol) in absolute dioxane (20 mL) was stirred vigorously, treated with diisopropylcarbodiimide (0.52 mL, 3.3 mmol), and stirred for 2 h (course of reaction monitored by TLC). The resulting activated ester was treated with cytosine (0.63 g, 3.3 mmol) and stirred vigorously for 2-4 h (course of reaction monitored by TLC). When the reaction was complete, water (200 mL) was added and acidified to pH 5-6. The resulting precipitate was filtered off and crystallized from propan-2-ol.

***N*-[(3,5)-Dimethyl-7-oxofuro[3,2-*g*]chromen-6-yl]acetyl]cytosine (13).** Yield 84%, C<sub>26</sub>H<sub>24</sub>N<sub>2</sub>O<sub>5</sub>, mp 276.5-278°C. PMR spectrum (300 MHz, DMSO-d<sub>6</sub>, δ, ppm, J/Hz): 1.99 and 2.08 (2H, 2 br.s, CH<sub>2</sub>-8), 2.27 (6H, s, CH<sub>3</sub>-3'', CH<sub>3</sub>-5''), 2.54 (1H, m, H-9), 2.85-3.06 (2H, m, H-7, H-11a), 3.38-3.85 (4H, m, CH<sub>2</sub>-10, CH<sub>2</sub>-13), 4.06 and 4.24 (2H, 2 d, J = 15.6, CH<sub>2</sub>-1'), 4.48 and 4.58 (1H, 2 d, J = 13.2, H-11e), 6.09 and 6.18 (1H, 2 d, J = 6.9, H-5), 6.22 (1H, d, J = 6.9, H-3), 7.24-7.32 (1H, m, H-4), 7.46 (1H, s, H-9''), 7.74 (1H, br.s, H-4''), 7.86 (1H, br.s, H-2'').

***N*-[(3,5,9)-Trimethyl-7-oxofuro[3,2-*g*]chromen-6-yl]acetyl]cytosine (14).** Yield 79%, C<sub>27</sub>H<sub>26</sub>N<sub>2</sub>O<sub>5</sub>, mp 307.5-309°C. PMR spectrum (300 MHz, DMSO-d<sub>6</sub>, δ, ppm, J/Hz): 1.97 and 2.09 (2H, 2 br.s, CH<sub>2</sub>-8), 2.25 (3H, s, CH<sub>3</sub>-3''), 2.46 (6H, s, CH<sub>3</sub>-5'', CH<sub>3</sub>-9''), 2.54 (1H, m, H-9), 2.80-3.02 (2H, m, H-7, H-11a), 3.40-3.86 (4H, m, CH<sub>2</sub>-10, CH<sub>2</sub>-13), 4.09 and 4.28 (2H, 2 d, J = 15.6, CH<sub>2</sub>-1'), 4.44 and 4.58 (1H, 2 d, J = 13.2, H-11e), 6.09 and 6.21 (1H, 2 d, J = 6.9, H-5), 6.22 (1H, d, J = 6.9, H-3), 7.24-7.34 (1H, m, H-4), 7.68 (1H, s, H-2''), 7.74 (1H, s, H-4'').

***N*-[(2,3,5)-Trimethyl-7-oxofuro[3,2-*g*]chromen-6-yl]acetyl]cytosine (15).** Yield 86%, C<sub>27</sub>H<sub>26</sub>N<sub>2</sub>O<sub>5</sub>, mp 293-294.5°C. PMR spectrum (300 MHz, DMSO-d<sub>6</sub>, δ, ppm, J/Hz): 1.95 and 2.04 (2H, 2 br. s, CH<sub>2</sub>-8), 2.16 (3H, s, CH<sub>3</sub>-3''), 2.36 (3H, s, CH<sub>3</sub>-2''), 2.39 (3H, s, CH<sub>3</sub>-5''), 2.54 (1H, m, H-9), 2.84-2.98 (2H, m, H-7, H-11a), 3.39-3.86 (4H, m, CH<sub>2</sub>-10, CH<sub>2</sub>-13), 4.07 and 4.25 (2H, 2 d, J = 15.6, CH<sub>2</sub>-1'), 4.42 and 4.59 (1H, 2 d, J = 13.2, H-11e), 6.10 and 6.19 (1H, 2 d, J = 6.9, H-5), 6.22 (1H, d, J = 6.9, H-3), 7.27-7.32 (1H, m, H-4), 7.35 (1H, s, H-9''), 7.67 and 7.69 (1H, 2 s, H-4'').

***N*-[(2,3,5,9)-Tetramethyl-7-oxofuro[3,2-*g*]chromen-6-yl]acetyl]cytosine (16).** Yield 86%, C<sub>28</sub>H<sub>28</sub>N<sub>2</sub>O<sub>5</sub>, mp 318-319.5°C.

PMR spectrum (300 MHz, DMSO-d<sub>6</sub>, δ, ppm, J/Hz): 1.95 and 2.04 (2H, 2 br. s, CH<sub>2</sub>-8), 2.16 (3H, s, CH<sub>3</sub>-3''), 2.39 and 2.43 (9H, 2 s, CH<sub>3</sub>-2'', CH<sub>3</sub>-5'', CH<sub>3</sub>-9''), 2.54 (1H, m, H-9), 2.82-3.02 (2H, m, H-7, H-11a), 3.40-3.85 (4H, m, CH<sub>2</sub>-10, CH<sub>2</sub>-13), 4.09 and 4.26 (2H, 2 d, J = 15.6, CH<sub>2</sub>-1'), 4.39 and 4.58 (1H, 2 d, J = 13.2, H-11e), 6.10 and 6.19 (1H, 2 d, J = 6.9, H-5), 6.22 (1H, d, J = 6.9, H-3), 7.28 (1H, t, J = 6.9, H-4), 7.52 (1H, s, H-4'').

***N*-[(4-Methyl-2-oxo-6,7,8,9-tetrahydro[1]benzofuro[3,2-*g*]chromen-3-yl]acetyl]cytosine (17).** Yield 84%, C<sub>29</sub>H<sub>28</sub>N<sub>2</sub>O<sub>5</sub>, mp 200.5-202°C.

PMR spectrum (300 MHz, DMSO-d<sub>6</sub>, δ, ppm, J/Hz): 1.85 (2H, m, CH<sub>2</sub>-7''), 1.92 (2H, m, CH<sub>2</sub>-8''), 1.96 and 2.08 (2H, 2 br. s, CH<sub>2</sub>-8), 2.46 (3H, s, CH<sub>3</sub>-4''), 2.54 (1H, m, H-9), 2.63 (2H, m, CH<sub>2</sub>-6''), 2.73 (2H, m, CH<sub>2</sub>-9''), 2.84-3.02 (2H, m, H-7, H-11a), 3.38-3.84 (4H, m, CH<sub>2</sub>-10, CH<sub>2</sub>-13), 4.09 and 4.27 (2H, 2 d, J = 15.6, CH<sub>2</sub>-1'), 4.40 and 4.57 (1H, 2 d, J = 13.2, H-11e), 6.10 and 6.20 (1H, 2 d, J = 6.9, H-5), 6.23 (1H, d, J = 6.9, H-3), 7.28 (1H, t, J = 6.9, H-4), 7.41 (1H, s, H-11''), 7.71 (1H, br. s, H-5'').

***N*-[4,11-Dimethyl-2-oxo-6,7,8,9-tetrahydro[1]benzofuro[3,2-*g*]chromen-3-yl]acetyl]cytosine (18).** Yield 81%, C<sub>30</sub>H<sub>30</sub>N<sub>2</sub>O<sub>5</sub>, mp 287-289°C.

PMR spectrum (300 MHz, DMSO-*d*<sub>6</sub>, δ, ppm, J/Hz): 1.84 (2H, m, CH<sub>2</sub>-7''), 1.93 (2H, m, CH<sub>2</sub>-8''), 1.94 and 2.04 (2H, 2 br. s, CH<sub>2</sub>-8), 2.44 (3H, s, CH<sub>3</sub>-4'', CH<sub>3</sub>-11''), 2.54 (1H, m, H-9), 2.61 (2H, m, CH<sub>2</sub>-6''), 2.73 (2H, m, CH<sub>2</sub>-9''), 2.85-3.01 (2H, m, H-7, H-11a), 3.39-3.82 (4H, m, CH<sub>2</sub>-10, CH<sub>2</sub>-13), 4.08 and 4.25 (2H, 2 d, J = 15.6, CH<sub>2</sub>-1'), 4.41 and 4.58 (1H, 2 d, J = 13.2, H-11e), 6.12 and 6.21 (1H, 2 d, J = 6.9, H-5), 6.22 (1H, d, J = 6.9, H-3), 7.27 (1H, t, J = 6.9, H-4), 7.52 (1H, br. s, H-5'').

***N*-[3-(3,5-Dimethyl-7-oxofuro[3,2-*g*]chromen-6-yl)propionyl]cytosine (19).** Yield 87%, C<sub>27</sub>H<sub>26</sub>N<sub>2</sub>O<sub>5</sub>, mp 185.5-187°C.

PMR spectrum (300 MHz, DMSO-*d*<sub>6</sub>, δ, ppm, J/Hz): 1.95-2.04 (2H, m, CH<sub>2</sub>-8), 2.29 (3H, s, CH<sub>3</sub>-3''), 2.39 (3H, s, CH<sub>3</sub>-5''), 2.20-2.75 (5H, m, H-9, CH<sub>2</sub>-1', CH<sub>2</sub>-2'), 2.82 (1H, m, H-11a), 3.14 (1H, m, H-7), 3.34 and 3.42 (1H, 2 d, J = 13.2, H-13a), 3.60-3.70 (1H, m, H-10a), 3.82-4.09 (2H, m, H-10e, H-13e), 4.42 and 4.64 (1H, 2 d, J = 13.2, H-11e), 6.12-6.21 (2H, m, H-3, H-5), 7.20-7.28 (1H, m, H-4), 7.48 (1H, br. s, H-9''), 7.76 (1H, s, H-4''), 7.90 (1H, br. s, H-2'').

***N*-[3-(3,5,9-Trimethyl-7-oxofuro[3,2-*g*]chromen-6-yl)propionyl]cytosine (20).** Yield 69%, C<sub>28</sub>H<sub>28</sub>N<sub>2</sub>O<sub>5</sub>, mp 153.5-155°C.

PMR spectrum (300 MHz, DMSO-*d*<sub>6</sub>, δ, ppm, J/Hz): 1.94-2.05 (2H, m, CH<sub>2</sub>-8), 2.25 (3H, s, CH<sub>3</sub>-3''), 2.38 (3H, s, CH<sub>3</sub>-9''), 2.44 (3H, s, CH<sub>3</sub>-5''), 2.20-2.75 (5H, m, H-9, CH<sub>2</sub>-1', CH<sub>2</sub>-2'), 2.82 (1H, m, H-11a), 3.14 (1H, m, H-7), 3.34 and 3.42 (1H, 2 d, J = 13.2, H-13a), 3.60-3.70 (1H, m, H-10a), 3.82-4.09 (2H, m, H-10e, H-13e), 4.42 and 4.64 (1H, 2 d, J = 13.2, H-11e), 6.12-6.21 (2H, m, H-3, H-5), 7.20-7.28 (1H, m, H-4), 7.65 (1H, s, H-2''), 7.74 (1H, s, H-4'').

***N*-[3-(2,3,5-Trimethyl-7-oxofuro[3,2-*g*]chromen-6-yl)propionyl]cytosine (21).** Yield 78%, C<sub>28</sub>H<sub>28</sub>N<sub>2</sub>O<sub>5</sub>, mp 221.5-223°C.

PMR spectrum (300 MHz, DMSO-*d*<sub>6</sub>, δ, ppm, J/Hz): 1.90-2.05 (2H, m, CH<sub>2</sub>-8), 2.19 (3H, s, CH<sub>3</sub>-3''), 2.36 (3H, s, CH<sub>3</sub>-2''), 2.39 (3H, s, CH<sub>3</sub>-5''), 2.20-2.69 (5H, m, H-9, CH<sub>2</sub>-1', CH<sub>2</sub>-2'), 2.82 (1H, m, H-11a), 3.14 (1H, m, H-7), 3.32 and 3.43 (1H, 2 d, J = 13.2, H-13a), 3.60-3.70 (1H, m, H-10a), 3.83-4.09 (2H, m, H-10e, H-13e), 4.43 and 4.62 (1H, 2 d, J = 13.2, H-11e), 6.08-6.19 (2H, m, H-3, H-5), 7.20-7.30 (1H, m, H-4), 7.34 and 7.38 (1H, 2 d, H-9''), 7.70 (1H, s, H-4'').

***N*-[3-(2,3,5,9-Tetramethyl-7-oxofuro[3,2-*g*]chromen-6-yl)propionyl]cytosine (22).** Yield 87%, C<sub>29</sub>H<sub>30</sub>N<sub>2</sub>O<sub>5</sub>, mp 146-147.5°C.

PMR spectrum (300 MHz, DMSO-*d*<sub>6</sub>, δ, ppm, J/Hz): 1.86-2.04 (2H, m, CH<sub>2</sub>-8), 2.16 (3H, s, CH<sub>3</sub>-3''), 2.33 (3H, s, CH<sub>3</sub>-2''), 2.42 (6H, s, CH<sub>3</sub>-5'', CH<sub>3</sub>-9''), 2.20-2.70 (5H, m, H-9, CH<sub>2</sub>-1', CH<sub>2</sub>-2'), 2.81 (1H, m, H-11a), 3.14 (1H, m, H-7), 3.30 and 3.39 (1H, 2 d, J = 13.2, H-13a), 3.58-3.70 (1H, m, H-10a), 3.83-4.10 (2H, m, H-10e, H-13e), 4.42 and 4.64 (1H, 2 d, J = 13.2, H-11e), 6.05-6.16 (2H, m, H-3, H-5), 7.20-7.28 (1H, m, H-4), 7.50 (1H, s, H-4'').

***N*-[3-(4-Methyl-2-oxo-6,7,8,9-tetrahydro[1]benzofuro[3,2-*g*]chromen-3-yl)propionyl]cytosine (23).** Yield 73%, C<sub>30</sub>H<sub>30</sub>N<sub>2</sub>O<sub>5</sub>, mp 239.5-241°C.

PMR spectrum (300 MHz, DMSO-*d*<sub>6</sub>, δ, ppm, J/Hz): 1.80-2.05 (6H, m, CH<sub>2</sub>-8, CH<sub>2</sub>-7'', CH<sub>2</sub>-8''), 2.33 (3H, s, CH<sub>3</sub>-4''), 2.20-2.70 (7H, m, H-9, CH<sub>2</sub>-1', CH<sub>2</sub>-2', CH<sub>2</sub>-6''), 2.73 (2H, m, CH<sub>2</sub>-9''), 2.82 (1H, m, H-11a), 3.14 (1H, m, H-7), 3.31 and 3.40 (1H, 2 d, J = 13.2, H-13a), 3.60-3.70 (1H, m, H-10a), 3.84-4.09 (2H, m, H-10e, H-13e), 4.44 and 4.62 (1H, 2 d, J = 13.2, H-11e), 6.06-6.17 (2H, m, H-3, H-5), 7.20-7.28 (1H, m, H-4), 7.38 and 7.40 (1H, 2 s, H-9''), 7.68 (1H, s, H-5'').

***N*-[3-(4,11-Dimethyl-2-oxo-6,7,8,9-tetrahydro[1]benzofuro[3,2-*g*]chromen-3-yl)propionyl]cytosine (24).** Yield 86%, C<sub>31</sub>H<sub>32</sub>N<sub>2</sub>O<sub>5</sub>, mp 192-193.5°C.

PMR spectrum (300 MHz, DMSO-*d*<sub>6</sub>, δ, ppm, J/Hz): 1.80-2.05 (6H, m, CH<sub>2</sub>-8, CH<sub>2</sub>-7'', CH<sub>2</sub>-8''), 2.31 (3H, s, CH<sub>3</sub>-11''), 2.42 (3H, s, CH<sub>3</sub>-4''), 2.20-2.70 (7H, m, H-9, CH<sub>2</sub>-1', CH<sub>2</sub>-2', CH<sub>2</sub>-6''), 2.74 (2H, m, CH<sub>2</sub>-9''), 2.83 (1H, m, H-11a), 3.14 (1H, m, H-7), 3.30 and 3.39 (1H, 2 d, J = 13.2, H-13a), 3.58-3.66 (1H, m, H-10a), 3.83-4.08 (2H, m, H-10e, H-13e), 4.44 and 4.64 (1H, 2 d, J = 13.2, H-11e), 6.06-6.17 (2H, m, H-3, H-5), 7.20-7.28 (1H, m, H-4), 7.48 (1H, s, H-5'').

## ACKNOWLEDGMENT

We thank OAO Eximed (Kiev, Ukraine) for assistance with the work.

## REFERENCES

1. R. D. H. Murray, *Progress in the Chemistry of Organic Natural Products*, Vol. 83, The Naturally Occurring Coumarins, Springer, Vienna and New York (2002).
2. L. Santana, E. Uriarte, F. Roleira, N. Milhazes, and F. Borges, *Curr. Med. Chem.*, 3239 (2004).
3. A. M. Gazaliev, M. Zh. Zhurinov, and B. I. Tuleuov, *Khim. Prir. Soedin.*, 301 (1991).
4. A. M. Gazaliev, M. Zh. Zhurinov, and S. D. Fazylov, *New Bioactive Alkaloid Derivatives* [in Russian], Gylym, Alma-Ata (1992), pp. 88-133.
5. T. V. Khakimova, O. A. Pukhlyakova, G. A. Shevaleeva, A. A. Fatykhov, E. V. Vasil'eva, and L. V. Spirikhin, *Khim. Prir. Soedin.*, 301 (2001).
6. A. D. Grebenyuk, V. I. Vinogradova, and A. K. Tashmukhamedova, *Khim. Prir. Soedin.*, 151 (2002).
7. K. A. Krasnov, V. G. Kartsev, A. S. Gorovoi, and V. N. Khrustalev, *Khim. Prir. Soedin.*, 367 (2002).
8. V. A. Saprykina, V. I. Vinogradova, R. F. Ambartsumova, T. F. Ibragimov, A. Sultankulov, and Kh. M. Shakhidoyatov, *Khim. Prir. Soedin.*, 479 (2004).
9. I. V. Nagorichna, I. P. Dubovik, M. M. Garazd, and V. P. Khilya, *Khim. Prir. Soedin.*, 196 (2003).
10. M. M. Garazd, Ya. L. Garazd, A. S. Ogorodniichuk, and V. P. Khilya, *Bioorg. Khim.*, 324 (2004).
11. A. A. Gershkovich and V. K. Kibirev, *Chemical Synthesis of Peptides* [in Russian], Naukova Dumka, Kiev (1992), p. 71.
12. G. W. Anderson, J. Zimmerman, and F. M. Callahan, *J. Am. Chem. Soc.*, **85**, 3039 (1963).